

Hair Growth Promoting Activity of *Nothopanax scutellarium* Merr. Leaves

Via Rifkia¹, Mahdi Jufri¹, Abdul Mun'im^{2*}

¹Graduate Program of Herbal Medicine, Faculty of Pharmacy, Universitas Indonesia, Depok 16424, West Java, INDONESIA.

²Department of Pharmacognosy-Phytochemistry, Faculty of Pharmacy Universitas Indonesia, Depok 16424, West Java, INDONESIA.

ABSTRACT

Objective: The aims of this study were to know the safety using Hen's eggs-chorioallantoic membrane (HET-CAM) test method and hair growth promoting activity of the ethyl acetate fraction of *Nothopanax scutellarium* leaves. **Methods:** Safety test was measured by scoring and categorizing irritation on HET-CAM. Meanwhile, activity test was conducted by applying the hair tonic of the ethylacetate fraction on the back of the rabbits, and the length of hair was measured in the 1st, 2nd and 3rd week. In the 3rd week, the hair growth was weighed and hair diameter also was measured using scanning electron microscope (SEM). **Results:** The result showed that 0.2 gram of ethyl acetate fraction of *N. scutellarium* leaves have mild irritation effect, whereas the formulation contained with 0.5% and 1% of fraction increased hair growth and hair diameter. **Conclusion:** The ethyl acetate fraction of *N. scutellarium*

have mild irritant effect, and the hair tonic demonstrated hair growth promoting activity.

Key words: Hair Tonic, Hair Growth Activity, *Nothopanax scutellarium*, Safety, HET-CAM.

Correspondence :

Abdul Mun'im, Department of Pharmacognosy-Phytochemistry, Faculty of Pharmacy Universitas Indonesia, Depok 16424, West Java, INDONESIA.

Phone no: +62 8111184550

Email: abdul.munim61@ui.ac.id

DOI: 10.5530/jyp.2017.9.85

INTRODUCTION

Hair loss and thinning are common problems that found in term of clinical dermatology. There are a number of products of hair tonic that claimed to increase hair growth, either using natural products or synthetic compounds. Some studies also reported the prospect of the natural product for hair growth promoting.¹⁻³ There has been no report on its main active ingredient responsible for the hair growth activity. In the current work, cedrol as a major constituent from *P. orientalis* was evaluated for its potential on hair growth *in vivo*, different concentration of cedrol (10, 20 and 30 mg/mL) promoted hair growth in a dose-dependent manner.² *Nothopanax scutellarium* Merr from Araliace family is the tropical plant. The leaves were traditionally known to have hair growth promotion activity.⁴

Based on previous research, ethanolic extract of the leaves was proven to have hair growth activity. However, the results of studies showed that hair growth activity of the ethanolic extract was lower than Minoxidil.⁵ Another study, the combination of ethanolic extract of the leaves with ethanol extract of tea leaves, also resulted in lower hair growth activity than positive control.⁶ Some studies reported that flavonoid demonstrated hair growth promoting activity.^{7,8,9} To increase hair growth activity of leaves extracts from *N. scutellarium*, further fractionation process was needed using the suitable solvent to increase the level of flavonoids, which required in increasing hair growth. Ethyl acetate was chosen as a solvent for a fractionation process of *N. scutellarium* leaves.

This study presented the safety test of the ethyl acetate fraction of *N. scutellarium* leaves using HET-CAM (Hen's Egg Test - chorioallantoic membrane) method. Then, the hair tonic formula of the fraction on hair growth activity was performed in rabbits.

MATERIALS AND METHODS

Materials

The leaves were obtained from the Indonesian Spice and Medicinal Crops Research Institute (ISMCRI), Bogor, and authenticated in Botany Herbarium Research Institute, Cibinong, West Java. The voucher specimen was deposited in Herbarium of Pharmacognosy, Faculty of Pharmacy, Universitas Indonesia. Solvents (ethanol, ethyl acetate, n-hexane), nipagin, nipasol, sodium metabisulfite, propylene glycol, chorioallantoic membrane (CAM), and minoxidil as the positive control were purchased from local suppliers.

Extraction and Fractionation

The leaves powder (4 kg) were macerated with ethanol (40L). Solvents were filtered and concentrated using rotary vacuum evaporator at 50°C. The ethanolic extract was dispersed in water and partitioned with n-hexane and ethyl acetate, subsequently. The ethyl acetate phase was concentrated using a rotary vacuum evaporator at a temperature of 50°C to give a viscous fraction of ethyl acetate. Total flavonoid of the extract was determined by AlCl₃ according to Kabir et al, 2016 with slight modification.^{10,11}

Safety test by HET-CAM method

The HET-CAM bioassay was performed to evaluate the level of irritation on mucous membrane based on slight modification of Steiling method.¹² Briefly, the fraction (0.2 g) solution was applied to chorioallantoic membrane (CAM) and left for 20 seconds until the sample was spread smooth. Then, the membrane was evaluated within 5 minutes to notice any symptoms of hemorrhage, lysis, and/or coagulation. The evaluation determined based on the result of scores and category of irritation (Table 1). The results of irritation scores can be calculated using the following equation:

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

where:

Hemorrhage time = time (in seconds) of the first appearance of blood hemorrhages.

Lysis time = time (in seconds) of the first appearance of vessel lysis.

Coagulation time = time (in seconds) of the first appearance of protein coagulation.

Formulation of hair tonic

Formulation of hair tonic prepared the variation of concentration of ethyl acetate fraction the leaves. The calculation of the percentage composition of each hair tonic can be seen in Table 2. Nipagin and nipasol were dissolved in ethanol, while sodium metabisulfite was dissolved in distilled water. Both solutions were mixed, and ethyl acetate fraction of *N. scutellarium* leaves was added into solution. Propylene glycol was added until the desired volume. All material were stirred until homogeneously mixed.

Evaluation of hair tonic

Evaluation of hair tonic preparations was conducted by observing the color and odor during storage. Furthermore, examination of pH by using a pH meter and hair tonic stability test at low temperature ($4^{\circ}\pm 2^{\circ}\text{C}$), room temperature ($25^{\circ}\pm 2^{\circ}\text{C}$), and high temperature ($40^{\circ}\pm 2^{\circ}\text{C}$) for 12 weeks by observing organoleptic appearance and pH every 2 weeks.^{15,16}

Hair growth activity

White male rabbits of New Zealand strain (4-5 months, with weights ranging from 2-4 kg) were obtained from Indonesia Research Institute of Animal Production (IRIAP). The study was approved by the Ethical Committee, Faculty of Medicine, Universitas Indonesia (No.683/H2.F1/ETIK/2012).

Four white male rabbits were used to examine hair growth activity. The hair on the back of the rabbit was shaved and divided into 6 areas with size of 4 x 4 cm; 3 areas on the left side, and another 3 areas on the right side with 2 cm distance of each area. Table 3 showed the treatment of samples. Hair tonic was applied in the shaved areas and observed for 3 weeks. During the experiment the hair growth was observed at week 1, 2, and 3, the 10 longest rabbit hair removed in each test area and measured using calipers.¹⁷ At the end of the application hair weight of rabbits were determined by pulling hair that grows on the test area and then weighed. The hair diameter was observed at week 3 by using SEM (Scanning Microscope Electron).

Statistical analysis

Data were reported as mean \pm standard deviation. Statistical analyses were performed using Student's t-test for the significance of the results ($P < 0.05$).

RESULTS

Fractionation

The result of fractionation that used in the preparation of hair tonic was ethyl acetate fraction. Characteristics of ethyl acetate fraction are shown in Table 4. The fraction contained total flavonoid 4.79%.

Safety test by HET-CAM method

Table showed the results of HET-CAM test. The irritation score of the test fraction is 4.54, indicated mild irritation on chorioallantoic membranes (Table 5).

Table 1: Scores and category of irritation

Scores on HET-CAM	Category of irritation
0-0.9	Nonirritant or practically no irritation
1-4.9	Weak or slight irritation
5-8.9	Moderate irritation
9-21	Strong or severe irritation

Table 2: The percentage of hair tonic ingredients

Ingredients	Concentration (%) (w/w)				
	Negative control (%)	Formula I (%)	Formula II (%)	Formula III (%)	Positive control (%)
The ethyl acetate fraction	-	0.25	0.5	1	-
Minoxidil	-	-	-	-	2
50% ethanol	60	60	60	60	60
Propylene glycol	2	2	2	2	2
Sodium metabisulfite	0.02	0.02	0.02	0.02	0.02
Nipagin	0.01	0.01	0.01	0.01	0.01
Nipasol	0.02	0.02	0.02	0.02	0.02
Aquadest	37.95	37.70	37.45	36.95	35.95

Table 3: Treatment of rabbits

Area	Group	Treatment
I	Normal control	Saline solution
II	Negative control	Hair tonic preparations containing no the ethyl acetate fraction
III	Formula I	Hair tonic preparations containing 0.25% the fraction
IV	Formula II	Hair tonic preparations containing 0.5% the fraction
V	Formula III	Hair tonic preparations containing 1% the fraction
VI	Positive control	Hair tonic preparations containing 2% minoxidil

Table 4: Characteristics of the ethyl acetate fraction

Characteristics of ethyl acetate fraction	
Organoleptic	Shape: Thick
	Color: Dark green
	Odor : Odorless typical

Evaluation of hair tonic

On this study, the stability test conducted by accelerated stability test. The results of organoleptic consisted of observations color and odor of hair tonic of the ethyl acetate fraction stored at low temperature ($4^{\circ}\pm 2^{\circ}\text{C}$), room temperature ($25^{\circ}\pm 2^{\circ}\text{C}$), and high temperature ($40^{\circ}\pm 2^{\circ}\text{C}$) looks stable. Results of hair tonic pH measurement of the fraction of ethyl acetate did not change significantly at low temperatures ($4^{\circ}\pm 2^{\circ}\text{C}$), room temperature ($25^{\circ}\pm 2^{\circ}\text{C}$) and high temperature ($40^{\circ}\pm 2^{\circ}\text{C}$) for 12 weeks of storage.

Effectivity of hair growth

The results showed that there were significant differences among groups. After observation for 3 weeks, the resulting hair growth on the groups of formula II containing fraction of test of 0.5% and formula III containing the fraction of test of 1%; has faster hair growth activity (Figure 1).

Hair length

The result of measurement of hair length of rabbit at week 1 shows the average on the groups of normal control, negative control, the formula I, formula II, formula III, and positive control respectively are 4.72 mm, 4.80 mm, 8.39 mm, 10.48 mm, 12.37 mm, and 9.16 mm. Furthermore, the results of the statistical analysis showed that the average hair length rabbit of each treatment group there were significant differences ($p < 0.05$).

At week 2, the average of hair length rabbit on the groups of normal control, negative control, the formula I, formula II, formula III, and positive control respectively are 8.10 mm, 8.13 mm, 12.09 mm, 14.29 mm, 18.24 mm, and 13.78 mm. The results of statistical calculation show the average hair length rabbit of each treatment group there were significant differences ($p < 0.05$). At week 3, the average of hair length rabbit on the groups of normal control, negative control, formula I, formula II, formula III, and positive control respectively are 12.33 mm, 12.35 mm, 16.25 mm, 18.36 mm, 28.25 mm, and 17.24 mm. The result of statistical calculation shows the average hair length rabbit of each treatment group there were significant differences ($p < 0.05$). Based on results of calculation of average hair length rabbits show formula II and formula III has faster activity of hair growth than the positive control (Figure 1). The re-

sult of statistical calculation shows the average hair length rabbit of each treatment group there were significant differences ($p < 0.05$).

Hair weight

The result of average of hair weight at normal control group, the negative control, formula I, formula II, formula III, and positive control each is 56.02 mg/cm², 55.86 mg/cm², 70.28 mg/cm², 73.02 mg/cm², 84.25 mg/cm², and 71.18 mg/cm². The results of average hair weight indicated that that test area which is given formula II and formula III containing ethyl acetate fraction of *N. scutellarium* leaves 0.5% and 1% had a greater weight than positive control (Figure 2). There was significant difference in each treatment ($p < 0.05$).

Hair diameter

The results of average rabbit's hair diameter show that the formula II and formula III containing of 0.5% and 1% ethyl acetate fraction of *Nothopanax scutellarium* leaves, respectively, has a larger diameter than the positive control (Figure 3). In addition to the observation of hair diameter, morphology hair of rabbit also was observed by SEM. The results show that the morphology hair of rabbit in the groups of formula II and formula III has composed regularly of hair cuticle and overlap, which consists of layers of flat keratin (Figure 4).

DISCUSSION

In this study, fractionation is conducted to increase the levels of flavonoids of *Nothopanax scutellarium* leaves, which believed to have potential activity of hair loss and thinning treatment. Flavonoid has been reported to have some pharmacological activities, such as antioxidant, hepatoprotective and anticancer.^{18,19,20} Based on several studies, a compound that plays a role in hair growth activity are proanthocyanidin compounds and procyanidin.^{8,21} One of the mechanisms hair growth promoting through Endothelial nitric oxide synthase (eNOS). Daidzein, genistein, isorhamnetin, kaempferol, quercetin, naringenin, and pelargonidin inhibited iNOS protein and mRNA expression and also nitric oxide (NO) production.¹⁶ Myricetin, quercetin, baicalein and fisetin demonstrated hair growth activity via inhibition of the type I 5 α -reductase.²²

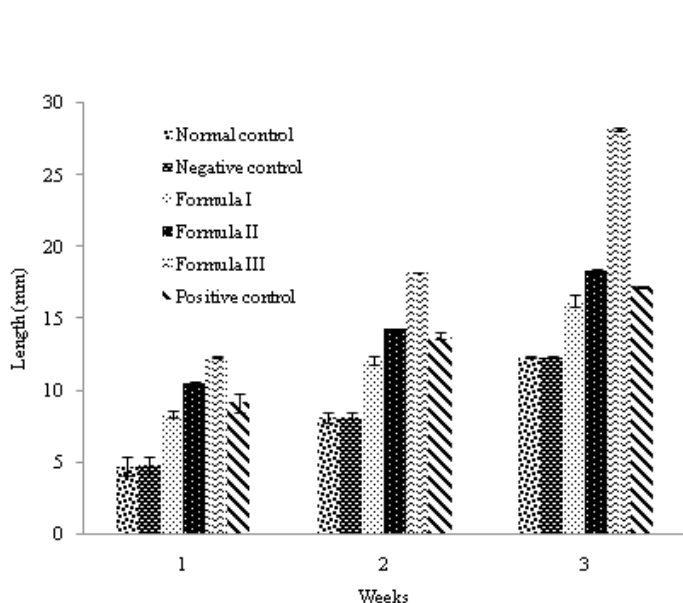


Figure 1: Effect of treatment on the average of hair length (mm).

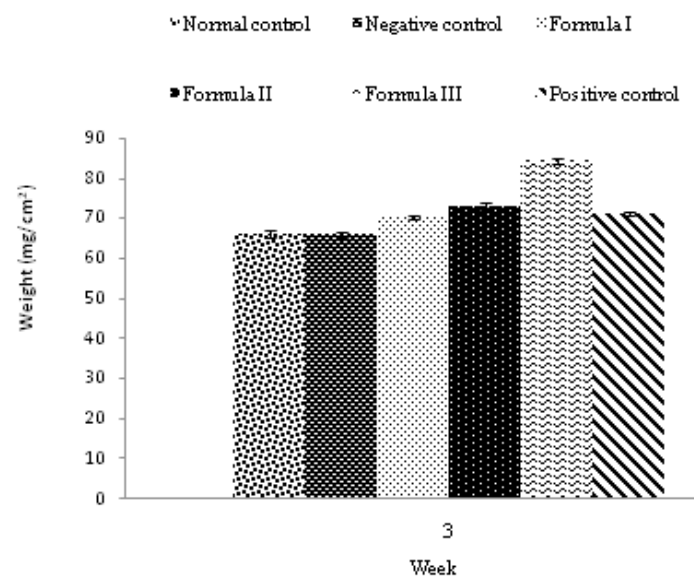
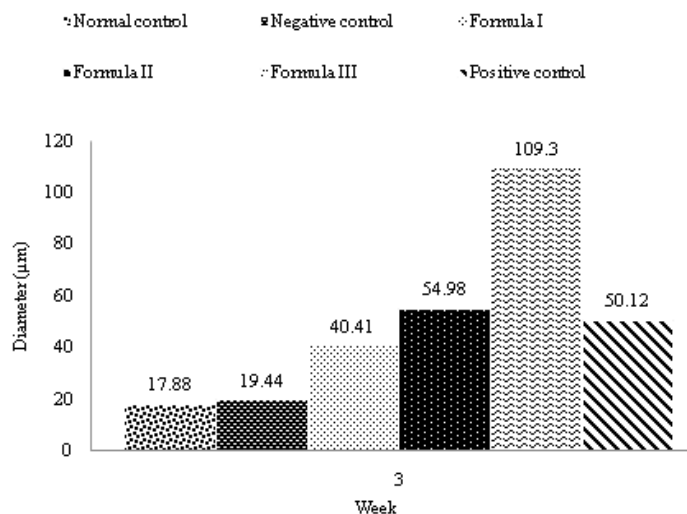
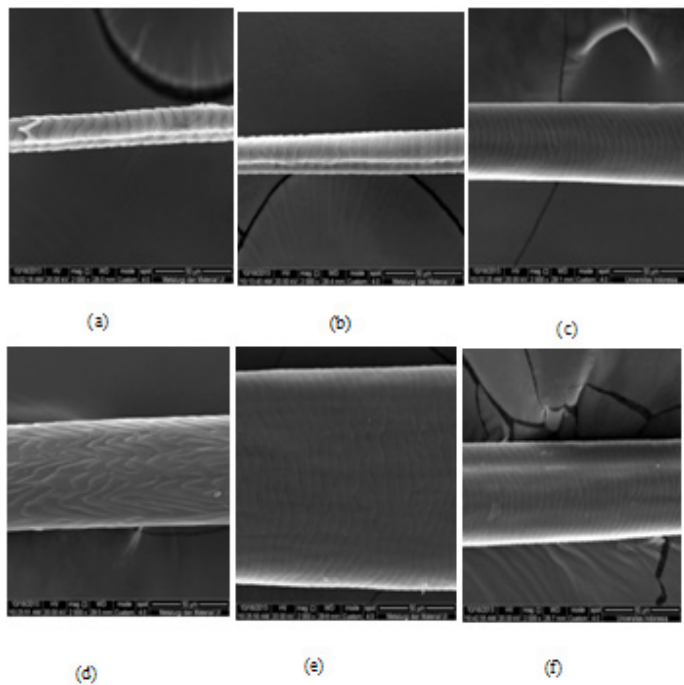


Figure 2: The average of hair weight (mg/cm²) of rabbit A, B, C, and D at the end of experiment.

Table 5: The result of average score and category of irritation

Sample	Average of Irritation score	Category of irritation
Negative control : 0.9%NaCl solution	0	Nonirritant or practically no irritation
Positive control : 1%SLS	10.23	Strong or severe irritation
Fraction of test: ethyl acetate fraction of <i>N. scutellarium</i> leaves	4.54	Weak or slight irritation

**Figure 3:** The average of rabbit hair diameter (µm) at the end of experiment.**Figure 4:** Scanning Microscope Electrone magnification: x2000. (a) normal control; (b) negative control; (c) formula I; (d) formula II; (e) formula III; (f) positive control.

Evaluation of irritation level was performed by using the HET-CAM method determined by the score and category irritation by comparing the changes in the chorioallantoic membrane between the negative control, positive control and fractions test. Results showed that there was a difference between the positive control (1%SLS) with test fractions containing ethyl acetate fraction of *N. scutellarium* leaves (Table 5). In some studies, testing by the HET-CAM method has been done especially for plants that can be developed into cosmetic products, such as plants of *Lansium domesticum* and *Phyllanthus niruri*. Both of these plants has efficacy as an antioxidant and antityronase. Based on the previous study, at a concentration of 2.5% either extract of *Lansium domesticum* or extract of *Phyllanthus niruri* cause moderate irritation of chorioallantoic membranes.²³

In this study, we used the ethyl acetate fraction as an active compound of the hair tonic formulation. This formulation was evaluated by observing the color, odor, pH, and stability of the hair tonic stored at low temperature ($4^{\circ}\pm 2^{\circ}\text{C}$), room temperature ($25^{\circ}\pm 2^{\circ}\text{C}$), and high temperature ($40^{\circ}\pm 2^{\circ}\text{C}$) for 12 weeks with a method of accelerated stability. The results showed hair tonic stable during storage. A product which stable in accelerated stability tests, that the that is stable in storage at room temperature for one year.²⁴ Meanwhile, result of observations showed a hair tonic pH has a pH in the range of physiological pH "acid mantle" the skin or also called pH balance, which ranges from 4.5 to 6.5, and still in the scalp pH range, i.e., between 4, 0 to 5.8.^{25,26}

Treatment of hair loss and thinning from natural product was widely needed. Proanthocyanidin from grape seeds stimulated the proliferation of hair follicle cells of mice *in vitro* and stimulate hair growth cycle of the telogen phase to anagen phase *in vivo*.⁸ Root extract of *Sophora flavescens* has been reported to stimulate hair growth.⁸⁻²⁷

In this study, formula II and III, each containing 0.5% and 1% of ethyl acetate fraction have faster hair growth than positive control (Figure 1). The mechanism of hair growth from the ethyl acetate fraction *N. scutellarium* leaves is suggested due to flavonoids. Based on several studies, flavonoids contribute to the activity of hair growth by strengthening the walls of capillaries in the hair follicles, as well as improve blood circulation to nourish hair follicles that can increase hair growth.⁷⁻⁹ Other studies also shown that flavonoids can shorten the telogen phase and can prolong the anagen phase. Based on research, minoxidil can prolong the anagen phase by extending the dermal papilla cell survival in the hair follicles by increasing proliferation and anti-apoptotic effects that can stimulate hair growth in the anagen phase.²⁸ The role of flavonoids on the activity of hair growth is caused by several factors hair growth, such as insulin-like growth factors-1 (IGF-1), vascular endothelial growth factors (VEGF), keratinocyte growth factors (KGF), and hepatocyte growth factors (HGF), these factors have the effect of stimulating hair growth.^{8,9-27}

In this study, the hair tonic on the formula III has a larger diameter than the positive control and has hair cuticle composed regularly and overlap (Figure 4). Some research suggests that healthy hair is observed with SEM (Scanning Electron Microscope) can be seen clearly has the hair cuticle smooth edges, patterns of cuticle layer to are arranged neatly to the inside of the hair seemed protected.^{25,29} Meanwhile, damaged hair has cuticle edges were chipped or loose. If the hair cuticle is almost completely peeled off, will make cortex layer is exposed, for example, the branched hair ends and hair fragile.²⁵

CONCLUSION

Based on the results of this research concluded that the ethyl acetate fraction of *Nothopanax scutellarium* leaves could mild irritate the membranes chorioallantois and hair tonic preparations with the concentra-

tion of 0.5% and 1% have hair growth activity the faster and can enlarge the diameter of the hair compared to the positive control.

ACKNOWLEDGEMENT

This work financially was supported by Directorate of Higher Education, Ministry of Technology Research and Higher Education, Republik Indonesia via Hibah PUPT 2015.

CONFLICT OF INTEREST

There is no conflict of interest.

REFERENCES

- Rathi V, Rathi JC, Tamizharasi S. Development and evaluation of polyherbal formulations for hair growth potential. *Pharmacognosy Research*. 2009;1(4):234.
- Zhang Y, Han L, Chen SS, Guan J, Qu FZ, *et al.* Hair growth promoting activity of cedrol isolated from the leaves of *Platyclusus orientalis*. *Biomedicine & Pharmacotherapy*. 2016;83:641-7. <https://doi.org/10.1016/j.biopha.2016.07.022>; PMID:27459121.
- Sun YN, Cui L, Li W, Yan XT, Yang SY, *et al.* Promotion effect of constituents from the root of *Polygonum multiflorum* on hair growth. *Bioorganic & medicinal chemistry letters*. 2013;23(17):4801-5. <https://doi.org/10.1016/j.bmcl.2013.06.098>; PMID:23896496.
- Heyne K. *Tumbuhan Berguna Indonesia*. Jakarta: Yayasan Sarana Wana; 1987. (in Bahasa)
- Handojo Y. Physical stability and hair growth promoting activity of *Nothopanax scutellarium* Merr leaves extract gel. 2011.
- Sholikhah N. Efek campuran ekstrak daun Teh (*Camellia sinensis* L.) dan daun Mangkokan (*Nothopanax scutellarium* Merr.) terhadap pertumbuhan rambut kelinci jantan. 2008. (in Bahasa)
- Kobayashi N, Suzuki R, Koide C, Suzuki T, Matsuda H, *et al.* Effect of leaves of *Ginkgo biloba* on hair regrowth in C3H strain mice. *Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan*. 1993;113(10):718-24. https://doi.org/10.1248/yakushi1947.113.10_718; PMID:8254481.
- Takahashi T, Kamiya T, Yokoo Y. Proanthocyanidins from grape seeds promote proliferation of mouse hair follicle cells *in vitro* and convert hair cycle *in vivo*. *ACTA DERMATOVENEREOLÓGICA-STOCKHOLM*. 1998;78:428-32.
- Awe EO, Makinde JM. The hair growth promoting effect of *Russelia equisetiformis* (Schlect&Chan). *Journal of Natural Products (India)*. 2009;2:70-3.
- Shah M, Kabir H, Hossain MM, Kabir I, Rahman M, *et al.* Phytochemical screening, antioxidant, thrombolytic, α -amylase inhibition and cyto toxic activities of ethanol extract of *Stuednera colocasiiifolia* K. Koch leaves. *J Young Pharm* 2016;8(4):391-7. <https://doi.org/10.5530/jyp.2016.4.15>.
- Mun'im A, Ramadhani F, Chaerani K, Amelia L, Arrahman A. Effects of gamma irradiation on microbiological, phytochemical content, antioxidant activity and inhibition of angiotensin converting enzyme (ACE) activity of *Peperomia pelucida* (L.) Kunth. *J Young Pharm* 2017;9(1):118-21.
- Steiling W, Bracher M, Courtellemont P, De Silva O. The HET-CAM, a useful *in vitro* assay for assessing the eye irritation properties of cosmetic formulations and ingredients. *Toxicology in vitro*. 1999;13(2):375-84. [https://doi.org/10.1016/S0887-2333\(98\)00091-5](https://doi.org/10.1016/S0887-2333(98)00091-5).
- Cazedeu EC, Carvalho FC, Fiorentino FA, Gremião MP, Salgado HR. Corrosi-tex®, BCOP and HET-CAM as alternative methods to animal experimentation. *Brazilian Journal of Pharmaceutical Sciences*. 2009;45(4):759-66. <https://doi.org/10.1590/S1984-82502009000400021>.
- Rajpal Deshmukh G, Hema Kumar K, Suresh Reddy PV, Srinivasa Rao B, Venkata Satish Kumar C. Evaluation of eye irritation potential of aqueous leaf extract of *Achyranthes aspera* by *in vitro* and *in vivo* method. *ISRN toxicology*. 2012. <https://doi.org/10.5402/2012/693489>; PMID:23724295 PMID:PMC3658502.
- ICH. Stability Testing of New Drug Substances and Products Q1A(R2). *Int Conf Harmon*. 2003:24.
- Harry RG. *Harry's Cosmeticology 8th ed.* (Rieger MM, ed.). New York: Chemical Publishing Co Inc; 2010.
- Yoon JI, Al-Reza SM, Kang SC. Hair growth promoting effect of *Zizyphus jujuba* essential oil. *Food and chemical toxicology*. 2010;48(5):1350-4. <https://doi.org/10.1016/j.fct.2010.02.036>; PMID:20206225.
- Andarwulan N, Kurniasih D, Apriady RA, Rahmat H, Roto AV, *et al.* Polyphenols, carotenoids, and ascorbic acid in underutilized medicinal vegetables. *Journal of Functional Foods*. 2012;4(1):339-47. <https://doi.org/10.1016/j.jff.2012.01.003>.
19. Yadav DK, Ali M, Ghosh AK, Kumar B. Isolation of flavonoid from *Abies web-biana* leaves and its activity. *Pharmacognosy Journal*. 2016;8(4).
20. Chen D, Landis-Pivowar KR, Chen MS, Dou QP. Inhibition of protease activity by the dietary flavonoid apigenin is associated with growth inhibition in cultured breast cancer cells and xenografts. *Breast Cancer Research*. 2007;9(6):R80. <https://doi.org/10.1186/bcr1797>; PMID:18300387 PMID:PMC2246179.
21. Kamimura A, Takahashi T. Procyanidin B-3, isolated from barley and identified as a hair-growth stimulant, has the potential to counteract inhibitory regulation by TGF- β 1. *Experimental dermatology*. 2002;11(6):532-41. <https://doi.org/10.1034/j.1600-0625.2002.110606.x>; PMID:12473061.
22. Herman A, Herman AP. Mechanism of action of herbs and their active constituents used in hair loss treatment. *Fitoterapia* 2016;114(6):18-25. <https://doi.org/10.1016/j.fitote.2016.08.008>; PMID:27552901.
23. Wih LVV, Ranti AR, Wasitaatmadja SM, Suryaningsih, Junardy FD M. Penelitian bahan pencerah dan pelembab kulit dari tanaman Indonesia. *Majalah Ilmu Ke-farmasian* 2009;4:1-8. (in Bahasa)
24. Ansel H. *Pengantar Bentuk Sediaan Farmasi (Trans)*. 4th ed. Jakarta: UI-Press, Jakarta; 1989.
25. Mitsui T. *New Cosmetic Science*. Amsterdam: 1st ed. Amsterdam; 1997.
26. Barel AO, Paye M MH. *Handbook of Cosmetic Science and Technology*. 4th ed. New York: Informa Healthcare; 2014.
27. Roh SS, Kim CD, Lee MH, Hwang SL, Rang MJ, *et al.* The hair growth promoting effect of *Sophora flavescens* extract and its molecular regulation. *Journal of dermatological science*. 2002;30(1):43-9. [https://doi.org/10.1016/S0923-1811\(02\)00060-9](https://doi.org/10.1016/S0923-1811(02)00060-9).
28. Han JH, Kwon OS, Chung JH, Cho KH, Eun HC, *et al.* Effect of minoxidil on proliferation and apoptosis in dermal papilla cells of human hair follicle. *Journal of dermatological science*. 2004;34(2):91-8. <https://doi.org/10.1016/j.jderm-sci.2004.01.002>; PMID:15033191.
29. Lim SN. A Study of Hair Damage by Magic Straight Perm. *Applied Microscopy*. 2012;42(3):129-35. <https://doi.org/10.9729/AM.2012.42.3.129>.

Article History: Submission Date : 09-02-2017; Revised Date : 04-05-2017; Acceptance Date : 28-05-2017.

Cite this article: Rifkia V, Jufri M, Mun'im A. Hair Growth Promoting Activity of *Nothopanax scutellarium* Merr. Leaves. *J Young Pharm*. 2017;9(3):436-40.